AMENDMENTS TO THE SPECIFICATION

Please amend the paragraph on page 24, line 18, to line 25, as follows:

Compounds having a quaternary amino group and a function capable of binding to other substances include, for example, FM3-25 (R), betaine, ~~cartinine~~ carnitine, tetramethylrhodamine cadaverine, and rhodamineX, or derivatives of these compounds. FM3-25 (R), tetramethylrhodamine cadaverine, or rhodamineX, or a derivative of these compounds, in particular, is preferable since it is a fluorescent substance and can be utilized as a detector (dyeing marker).

Please amend the paragraph on page 26, line 4, to line 13, as follows:

When the charge control agent of the present invention is used, the optimum pH of a sample solution should be appropriately selected. For example, the pKa of a carboxylic acid group is approximately 2-5, whereby the carboxylic acid group is negatively charged since a proton is removed therefrom when the pH of the sample solution ~~is in a range of 2-5 or higher~~ greater than the above pKa. Thus, in order to cause the charge control agent having a carboxylic acid group to react with the target particle while keeping a negative charge, it is preferable that the pH of the sample solution be set so as to be 4 or higher.

Please amend the paragraph on page 26, line 14, to line 18, as follows:

Similarly, in the case where a charge control agent having a phenol group or an alcohol group is used, since the pKa of the phenol group and the alcohol group is approximately 9-11, it is preferable that the pH be set so as to be ~~in a range of 9-11 or higher~~ greater than the above pKa in order to use the agent while keeping a negative charge.

Please amend the paragraph on page 26, line 19, to line 23, as follows:

In the case where a charge control agent having a tertiary amine group is used, since the pKa of the tertiary amine group is approximately 10-12, it is preferable that the agent be used at the pH smaller than the above pKa ~~equal to or smaller than 10-12~~ in order to use it while keeping a negative charge.

Please amend the paragraph on page 28, line 6, to line 18, as follows:

An analysis of a lymphocyte surface antigen, whose test is easy to perform, is known as a test for an immune function. It is known that acute stress increases the number and the activity of NK cells (natural killer cells), which are a type of lymphocyte, in the peripheral ~~deletion~~ blood and that chronic stress decreases the number and the activity thereof. For example, decreased activity of the NK cells has been reported among patients with depression (TOMONOBU Kawano (ed.) "Handbook of Stress Diagnosis", Medical Science International, January 1990, p11, table 2-2). Also, it has been reported that stress associated with examinations decreases the activity of the NK cells and production of interferon (TOMONOBU Kawano (ed.) "Handbook of Stress Diagnosis", Medical Science International, January 1990, p11, table 2-2).

Please amend the paragraph on page 44, line 2, to line15, as follows:

A platinum loopful of bacterial cell body was removed from a slant solid culture of ST, separately inoculated into a 200 ml conical flask containing tryptic soybean culture prepared according to a common procedure, and incubated in static culture at 37℃ for 16 hours in atmosphere of 95% CO2. Similarly, a culture solution of SM was prepared. Each 500 μ l of the resultant culture solution was transferred to a 1.5 ml sample tube, and 5 μ l of Cascade Blue-labeled anti-ST antibody solution (1 mg/ml) and 5 μ l of Cascade Blue-labeled anti-SM antibody solution (1 mg/ml) were added thereto and left for 30 minutes at 37℃ in a dark place, thereby bringing streptococci into contact with a labeled antibody for each streptococcus. Next, the fluorescent capillary electrophoresis was performed therefor under the following conditions.

Please amend the paragraph on page 45, line 3, to line25, as follows:

Also, the fluorescent capillary electrophoresis was similarly performed for a solution (control sample) generated by adding 5 μ l of a solution containing either of the above two Cascade Blue-labeled anti-streptococcus antibodies to 500 μ l of 100mM Tris-boric acid buffer, and for a solution (comparison sample) generated as follows: each of

ten types of bacteria including Brucella sp. strain KYM-1, Stenotrophomonas sp. strain KYM2, Acinetobactersp. strain KYM3, Commanonas sp. strain KYM4, Aureobacterium sp. strain KYM6, Cellulomonas sp. strain KYM7, Acinetobacterium sp. strain KYM8, and three types of Escherichia coli was dissolved in 500 μ l of 100mM Tris-boric acid buffer so as to be 10^7/ml, 5 μ l of a solution containing either of the above two Cascade Blue-labeled anti-streptococcus antibodies was added thereto, and the resultant solution was left for 15 minutes at room temperature in a dark place. As a result, as for the control sample containing only the Cascade Blue-labeled anti-streptococcus ~~anti-salmonella~~ antibody in the Tris-boric acid buffer, a peak of the free Cascade Blue-labeled anti-streptococcus ~~anti-salmonella~~ antibody was detected at substantially the same residence time as the electroosmotic flow, and no other peak was detected. As is the case with the control, as for the solution (comparison sample) containing ten types of bacteria, only a peak of the free Cascade Blue-labeled anti-streptococcus ~~anti-salmonella~~ antibody was detected.